Original Article

Calculating the Rate of Senescence From Mortality Data: An Analysis of Data From the ERA-EDTA Registry

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Abstract

The rate of senescence can be inferred from the acceleration by which mortality rates increase over age. Such a senescence rate is generally estimated from parameters of a mathematical model fitted to these mortality rates. However, such models have limitations and underlying assumptions. Notably, they do not fit mortality rates at young and old ages. Therefore, we developed a method to calculate senescence rates from the acceleration of mortality directly without modeling the mortality rates. We applied the different methods to age group–specific mortality data from the European Renal Association—European Dialysis and Transplant Association Registry, including patients with end-stage renal disease on dialysis, who are known to suffer from increased senescence rates (n = 302,455), and patients with a functioning kidney transplant (n = 74,490). From age 20 to 70, senescence rates were comparable when calculated with or without a model. However, when using non-modeled mortality rates, senescence rates were yielded at young and old ages that remained concealed when using modeled mortality rates. At young ages senescence rates were negative, while senescence rates declined at old ages. In conclusion, the rate of senescence can be calculated directly from non-modeled mortality rates, overcoming the disadvantages of an indirect estimation based on modeled mortality rates.

Key Words: Senescence rate—Acceleration of mortality—Senescence—Aging—Modeling—Gompertz model.

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Across populations and species, actuarial mortality rates exhibit different age patterns (1). Mortality rates can increase, be constant, or decrease over age. Demographers interpret increasing mortality rates at the population level as a manifestation of senescence at the organismal level. Likewise, they interpret constant or decreasing mortality rates as a manifestation of absent senescence (2–5). Senescence is a result of manifold biological mechanisms that lead to an increasing vulnerability to death. Although biologists have made strong effort at explaining and measuring senescence, the nature of these biological mechanisms remains unclear and a reliable biomarker of senescence is lacking (6,7). Moreover, it is continuously debated whether senescence and disease are distinct or related phenomena (4,8).

The rate of mortality can be regarded as a speed function: it expresses the number of deaths per unit of time comparably with the speed of a car that is expressed as the number of driven meters per unit of time. It follows that an increase in mortality rate corresponds with an acceleration of mortality, while a decrease in mortality rate corresponds with a deceleration of mortality, similar to the acceleration or deceleration of the car (9,10). As senescence is represented by an increase in mortality rate over age, the rate of senescence can be calculated from the acceleration of mortality and is expressed as the increase in mortality rate per year of age (9,11). As an advantage to both demographers and biologists, this approach requires neither any assumptions on the age pattern of accelerations and decelerations of mortality nor on the biological mechanisms underlying the process of senescence.

In this study, we describe and test a method to calculate the rate of senescence from age-specific mortality rates directly from non-modeled mortality rates. This method can be used at all ages and is free of biological assumptions. It is purely based on the definition of senescence as the increase in mortality rate over age and calculates the rate of senescence as the acceleration of mortality over age. We apply this method to mortality data of patients with end-stage renal disease who are either on dialysis therapy or have undergone kidney transplantation. Patients on dialysis therapy are biologically and clinically known to suffer from increased senescence rates (12–14). Mortality rates and senescence rates in patients with a functioning transplant are lower than in those on dialysis (15) and approach those of the general population (14).

**Methods**

**Study Population**

Data were provided by the Registry of the European Renal Association—European Dialysis and Transplant Association, which records the treatment and survival history of European patients receiving renal replacement therapy, either dialysis or kidney transplantation (16). Patients were included when renal replacement therapy was started during a period from 1985 through 2011. Follow-up ended on January 1, 2012. Individual patient data were available from 1985 for Austria, the French-speaking region of Belgium, Finland, Greece, Iceland, the Netherlands, Norway, and Scotland, from 1994 for the Flemish-speaking region of Belgium, from 1990 for Denmark, from 2006 for Romania, from 1991 for Sweden, and from 1997 for England, Wales, and Northern Ireland. In addition, individual patient data were available for several regions in France from 2008, for several regions in Spain, including data from 1985 for Andalusia, from 2002 for Castile and León, from 2005 for Extremadura, from 2007 for Galicia, and from 1992 for Valencia.

Mortality rates were calculated based on the follow-up data contributed by each individual patient, separated for follow-up during dialysis and follow-up with a functioning transplant. For patients on dialysis, follow-up started 6 months after initiation of dialysis therapy, to account for early treatment-related mortality, and lasted until death, transplantation, recovery of renal function, loss to follow-up, or censoring on January 1, 2012. For patients with a functioning transplant, follow-up started 6 months after transplantation, to account for acute surgery-related mortality, and lasted until death, transfer to dialysis due to transplant failure, loss to follow-up, or censoring at January 1, 2012. For both treatment groups, the age-specific mortality rates were derived by dividing the number of deaths by the years of follow-up per 5-year age group.

As a reference, mortality rates were also calculated for the general European population. Numbers of deaths and population sizes were derived from Eurostat for the countries and regions included in this study (17). For each 5-year age group, the number of deaths was divided by the population size, both summed for the countries and regions and the years during which the countries and regions contributed data. As data were mostly available up to the age of 100, we excluded mortality rates from that age onward.

**Estimating the Acceleration of Mortality Indirectly**

**From the Parameters of Modeled Mortality Rates**

To compare the use of non-modeled mortality rates with the use of modeled mortality rates, the mortality rates were modeled with the Gompertz model and senescence rates were estimated as previously described (14). Considering the applicability of the model, mortality data were included for the ages of 20–85 years (14,18). The Gompertz model is mathematically described as \( m(t) = e^{\alpha t}, \) where \( m(t) \) is the mortality rate at age \( t \) in years and \( \alpha \) and \( \gamma \) are model parameters. The minimal mortality rate at \( t = 0 \) is determined by \( \alpha \), the subsequent exponential increase by \( \gamma \). On a logarithmical scale, the model conforms to a straight line, described by \( \ln m(t) = \ln \alpha + \gamma t \). The slope of this line is determined by \( \gamma \), describes the acceleration of mortality on the logarithmical scale, and estimates the relative senescence rate. The derivative function of the Gompertz model describes the acceleration of mortality on an absolute scale, estimates the absolute senescence rate, and is mathematically described as \( m'(t) = m(t) e^{\gamma t}. \)

**Calculating the Acceleration of Mortality Directly From Non-modeled Mortality Rates**

The method proposed here calculates the absolute senescence rate directly from non-modeled mortality rates on an absolute scale. The method is based on the mathematical definition of a derivative function (19). In general, for a given function \( y = f(x) \), the derivative function is \( f'(x) = dy/dx \), where \( d \) denotes an infinitesimal change in \( y \) or \( x \). In the case of mortality rate \( m \) calculated for age \( t \), the notations \( y \) and \( x \) are replaced: \( m'(t) = dm/dt \). When we take \( d \) as small as possible, \( dt \) corresponds to the difference in age between two age groups and \( dm \) equals the difference in mortality rate between both age groups. Using this method, we calculate the rate at which the mortality rate changes, thus the acceleration of mortality, between two age groups on average. Applied, we calculated the senescence rate of an age group as the mortality rate of the following age group minus the mortality rate of the age group of interest, divided by...
the difference in age between both age groups, the latter constantly being five years because of the use of 5-year age groups.

We excluded mortality rates from the analyses when the number of person-years was less than 200 per 5-year age group. Patients on dialysis aged 100 years and older were excluded, corresponding to less than 0.01% of follow-up and 0.01% of deaths. Patients with a functioning transplant aged 90 years and older were excluded, corresponding to 0.02% of follow-up and 0.05% of deaths. Due to the nature of this method, senescence rates could not be calculated for the oldest age group. It was not necessary to exclude mortality data for the youngest age groups.

Statistical Analyses

The calculations of the age-specific mortality rates were performed using IBM SPSS Statistics 20. The Gompertz model was fitted to the mortality data using Stata/SE 12.1, as previously described (14).

Results

Table 1 provides the general characteristics of the study population of patients with end-stage renal disease on dialysis or with a functioning kidney transplant. As patients could successively undergo dialysis treatment and kidney transplantation, some patients contributed follow-up to both treatment groups; this was the case for 59,781 patients (18.8%) and 554,809 years of follow-up (41.5%). The maximum number of different treatment periods per patient was 13.

In Figure 1, the different methods to infer senescence rates from mortality rates are compared. For comparison with the method proposed here, we replicated the estimations of senescence rates from mortality data that were modeled by the Gompertz model, as described previously (14). Figure 1A and B show the modeled mortality rates over age on a logarithmical scale and the relative senescence rates estimated from the model’s parameters. According to this method, senescence rates were constant over age, lowest in patients on dialysis, intermediate in patients with a functioning transplant, and highest in the general population. Figure 1C and D show the modeled mortality rates over age on an absolute scale and the absolute senescence rates estimated from the model’s parameters. According to this method, senescence rates increased over age, were highest in patients on dialysis, intermediate in patients with a functioning transplant, and lowest in the general population.

Table 1. Characteristics of the Study Population of Patients With End-Stage Renal Disease

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Total</th>
<th>On Dialysis</th>
<th>With a Functioning Kidney Transplant</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total number of patients, N</td>
<td>317,168</td>
<td>302,455</td>
<td>74,490</td>
</tr>
<tr>
<td>Sex, % male</td>
<td>61.5</td>
<td>61.5</td>
<td>62.7</td>
</tr>
<tr>
<td>Age, median years (iqr)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>At first treatment</td>
<td>64.1 (51.0–73.7)</td>
<td>65.0 (52.4–74.1)</td>
<td>48.6 (36.9–58.6)</td>
</tr>
<tr>
<td>At death</td>
<td>72.2 (63.5–78.8)</td>
<td>73.1 (64.8–79.3)</td>
<td>63.5 (54.7–70.6)</td>
</tr>
<tr>
<td>Follow-upper patient, median years (iqr)</td>
<td>2.7 (1.1–3.6)</td>
<td>2.0 (0.9–3.8)</td>
<td>5.4 (2.3–9.8)</td>
</tr>
<tr>
<td>By number of contributed years of follow-up</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total years of follow-up</td>
<td>1,337,832</td>
<td>841,109 (62.9)</td>
<td>496,723 (37.1)</td>
</tr>
<tr>
<td>Sex, % male</td>
<td>60.6</td>
<td>59.8</td>
<td>62.0</td>
</tr>
</tbody>
</table>

Note: The total study population includes both patients on dialysis and patients with a functioning kidney transplant. As patients could successively undergo dialysis treatment and kidney transplantation, patients can be represented in both the group on dialysis and the group with a functioning transplant. iqr = inter-quartile range.
We compared the outcomes of different methods to determine senescence rates from mortality rates. Classically, mortality rates are modeled by the Gompertz model and subsequently presented on a logarithmical scale, rendering easily comparable straight lines over age of which the slopes are determined by a single parameter of the Gompertz model (Figure 1A). Senescence rates are derived from the increase in these lines and are thus fixed over age (Figure 1B) (2,20–22). Due to the logarithmical scale, this method estimates the senescence rate as the relative acceleration of mortality over age. However, it has been theoretically objected that the senescence rate should be defined as the absolute acceleration of mortality over age (11,23–25). According to this view, we have proposed to model mortality rates on an absolute scale (Figure 1C). Earlier, we have demonstrated that senescence rates can be more adequately derived from the absolute acceleration of mortality over age, using the derivative function of the Gompertz model (Figure 1D) (14). Contrary to the relative senescence rates, but in line with biological and clinical knowledge, the absolute senescence rates were highest in patients on dialysis and lowest in the general population. The principle of using the derivative function can be applied to any model of mortality over age.

Modeled mortality data are not always preferably used over crude mortality data. The ability of mathematical models to describe mortality is limited to a specific age range. The Gompertz model does not fit mortality rates at the youngest and oldest ages (18,26). This is explained by the fact that these models necessarily mold mortality data into a prescribed pattern. The Gompertz model assumes mortality rates to increase exponentially over age (26). The mortality rates at the youngest and oldest ages in patients on dialysis and patients with a functioning transplant deviated from an exponential increase over age, which became only apparent when assessing non-modeled mortality rates (Figure 1E). Models as the Gompertz model do not account for such deviating age patterns of mortality rates that may be valuable to measure, as will be discussed hereafter. Moreover, these models, which are by themselves of only mathematical nature, are mistakenly interpreted biologically (26–28). Apart from the Gompertz model, the Weibull model and the logistic model are often used without any of them superiorly fitting mortality curves over age. The choice of a model is consequently based on biological assumptions explaining the age pattern that is imposed on the rates by the model (26,29). As we have shown previously (14), such a biological meaning is generally attributed to the mathematical parameters of the Gompertz model, but has no empirical foundation and is biologically invalid (11,23–25). An alternative method has been proposed to calculate senescence rates from non-modeled mortality rates, but only on a logarithmical scale, thus rendering relative senescence rates (30,31).

Here we extended our earlier analyses by applying a method that circumvents the need to first model mortality rates for the calculation of senescence rates on an absolute scale (Figure 1E and F). From age 20 to approximately 70, the senescence rates calculated by this method were similar to those estimated by the derivative function of the Gompertz model. However, this method additionally offers the possibility to calculate senescence rates for young and old ages that are unaccounted for by the Gompertz model.
At young ages, the method proposed here disclosed negative senescence rates. Senescence rates were more negative in patients on dialysis than in patients with a functioning transplant and in the general population. Negative senescence rates at young ages have not been frequently addressed for humans, as changes in mortality are mainly studied from adolescence onward. For several non-human species it has been recognized that mortality rates decline during life, a finding which has been termed negative senescence (32). At the beginning of human life, negative senescence is universally observed as a decline in mortality rate caused by processes that are distinct from senescence. It may be the result of a reduction in children’s vulnerability during development and growth or of a reduction in a population’s vulnerability due to the early death of the frailest children and the selective survival of healthier children (33,34). In the cases of dialysis therapy (35) and kidney transplantation (36), development and growth are impaired in children, although the deficits are partly compensated at a later age. The negative senescence rates in both groups more likely arise from a sharp decline in mortality rate due to early death of the frailest patients and the selective survival of healthier patients. Particularly in the youngest patients, mortality rates are high due to the underlying renal disease, congenital disorders that are associated with pediatric renal disease, and the complications of dialysis therapy or transplantation (37–39). The more negative senescence rates in children on dialysis compared with children with a functioning transplant and the general population can be explained as they display higher mortality rates at the youngest ages with a subsequent sharper decline in mortality and improved growth. In addition, their negative senescence rates may be more pronounced due to a stronger effect of selective survival of the less frail patients: those selected for transplantation are relatively healthier (38), the survival probabilities of children admitted to dialysis vary more than those of children undergoing kidney transplantation and in the general population. Negative senescence early in life has received little attention of gerontologists, but interacts closely with senescence later in life (34). It would be interesting to include the age at which negative senescence rates turn into positive rates in studies on senescence. Comparable turning points in the age pattern of senescence rates have been studied, but only at older ages (40).

At old ages, the method proposed here uncovered declining senescence rates that are not accounted for by many models. The phenomenon of leveling mortality rates, and thus declining senescence rates, at old ages has been described in different populations (2,41). Heterogeneity in survival patterns due to individual differences in frailty is thought to bring about leveling mortality rates (42). In selected homogeneous populations, mortality rates continue to increase exponentially up to the oldest ages (43). The decline in senescence rates in patients on dialysis and patients with a functioning transplant can be explained by the selective survival to the older ages of the relatively healthier patients and by the selection of relatively healthier patients to undergo these therapies at older ages. That the senescence rates in patients with a functioning transplant declined at younger ages compared with patients on dialysis suggests that these selective processes are stronger in this group, probably because of the reluctance to perform kidney transplantation in older patients. Senescence rates in the general population did not decrease, probably because heterogeneity effects emerge at older ages in large populations with low senescence rates (2,41). The method proposed here can be applied to further study declines in senescence rates in populations at the oldest ages.

This study investigated methods that determine senescence rates at the population level. The senescence rate of an individual cannot be inferred directly from the senescence rate of the population to which he belongs (24). Still, the increased senescence rates in patients on dialysis can be attributed to biological and clinical mechanisms that have been observed and promote senescence in the individual patients (8,44). In patients with end-stage renal disease as well as in older people in the general population, podocytes lining the epithelium of glomeruli show signs of damage and dysfunction, resulting in glomerulosclerosis and reduced glomerular filtration (45). Renal dysfunction causes accumulation of mineral and uremic toxins, oxidative stress, and systemic inflammation, which are normally seen at old ages. These processes promote cellular senescence through DNA damage, mitochondrial dysfunction, and telomere shortening. Widespread cellular injury and dysfunction lead to increased risks and rates of atherosclerosis, cardiac disease, cancer, immune deficiency, cognitive impairment, sarcopenia, and osteoporosis. These disorders, together with dialysis therapy itself, again induce further hemodynamic and immunological disturbances and loss of renal function. Eventually, these disorders lead to increased mortality rates (12,13,45,46). The method proposed here can be applied to compare the senescence rates of different populations, to identify populations with increased or decreased senescence rates, and to evaluate the effects of interventions on the senescence rate.

The method presented here has several advantages. As it does not use modeled mortality rates, it can be applied to any mortality data, at all ages, and independent of species, geographic origin, calendar period, and birth cohort. As it does not mold the mortality rates into a prescribed pattern, it closely follows the crude mortality data, does not discard any mortality data, and is very sensitive to changes in the senescence rates, as illustrated earlier for young and old ages. Finally, as this method is free of assumptions about the mortality patterns over age or the biological determinants underlying these patterns, it can only be interpreted based on its mathematical meaning. It measures the acceleration of mortality over age and thereby, given the definition of senescence as the increase in mortality rate over age, describes the senescence rate.

The use of non-modeled mortality data for the calculation of the senescence rate also has disadvantages. First, substantial amounts of age-specific crude data are required to prevent high variability in the estimates due to measurement error. Although large cohorts were available in this study, the senescence curves that were calculated with the proposed method show more variability than those calculated with the Gompertz model. We excluded age groups with few observations; each of these comprised less than 25 person-years of follow-up. Each age group included in the study comprised more than 200 person-years of follow-up. It is difficult to determine a level above which variability is unacceptably high. We cannot distinguish whether variability has arisen from measurement error or real age-related effects, which are both suppressed by modeling. Much of the variability depends on the width of the age groups for which the mortality rates have been calculated. This width can be adjusted in this method to reduce the effect of data variability. This reduction in variability will probably render the senescence rates more similar to those calculated from modeled mortality data. Furthermore, this method can be extended with techniques to smoothen the senescence curves, but that again introduces modeling based on statistical or biological assumptions. Second, the senescence rate can only be determined for those age groups with available mortality data. Without a model of mortality over age, extrapolation to other ages is not possible.
In conclusion, this study shows how the absolute rate of senescence can be calculated directly from age-specific non-modeled mortality rates. The methodology is simple, sensitive to changes in mortality over age, free of limitations in its applicability, and does not require any biological interpretation of mathematical models.

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References
16. ERA-EDTA Registry. ERA-EDTA Registry Annual Report 2011. Amsterdam, the Netherlands: Department of Medical Informatics, Academic Medical Center Amsterdam; 2013.